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General meetings
Held at 7:30 pm on the fourth Friday of each month at the Newborough Uniting Church, Old Sale Road Newborough VIC 3825

Apuffball fungus *Scleroderma* sp. beginning to split and release its spores at College Creek in June 2017 (Photo: Margaret Rowe).

Upcoming events

**November general meeting:** Friday 24 November – Strzelecki Warm Temperate Rainforest – Martin O’Brien
**Excursion:** Saturday 25 November – Uralla Reserve. Meet 10am at the reserve.

**Bird Challenge Count:** Thursday 30 November – Sunday 3 December

**Christmas Party:** Saturday 9 December at John & Jan Tulloch’s place, 55 Lynne Avenue, Moe South.

**January general meeting:** Friday 12 January – Summer Members’ Night
**Excursion:** Saturday 13 January – Wildflower Walk in Baw Baw NP with Friends of Baw Baw
**Club Summer Camp:** Friday 2 – Tuesday 6 February 2018 at Mount Buffalo

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The Ties that Bind: Fungi in Terrestrial Ecosystems 26.05.2017

Dr McMullan-Fisher, a mycologist on the Fungimap Conservation and Biodiversity Subcommittee, began her talk by explaining the focus of Fungimap on the occurrence of Tea-tree Fingers *Hypocreopsis amplectens*, the only macrofungus listed under the Victorian *Flora and Fauna Guarantee Act 1988*. Records of Tea-tree Fingers in southern Victoria seem to be in decline and the call is out for amateur fungi sleuths to keep an eye out.

Tea-tree Fingers grows on another wood-decomposing fungus that seems to prefer dead tea-tree substrates.

Dr McMullan-Fisher emphasised the three "F's" of an ecosystem: flora, fauna and fungi. The interactions between the kingdoms of living things are vital for maintaining healthy and diverse ecosystems. Many of these interactions are complex and surprising.

*Photobionts* are the primary producers in an ecosystem that capture carbon dioxide and, by using sunlight, turn it into carbohydrates and proteins. Plants, algae, bryophytes, cyanobacteria and lichens are examples of photobionts. Lichens consist of a fungal component, or mycobiont, and a photobiont component.

*Cyanolichens* contain a cyanobacteria that can fix nitrogen directly from the atmosphere.

For the digestion of their plant diets, many *herbivores* rely on gut microbes such as bacteria, protozoans and fungi. Anaerobic chytrid fungi that live in the digestive tracts of ruminants enable fermentation to take place.

*Carnivores* prey on other animals for their nutrition and there is a group of fungi that have adapted to 'capturing' nematodes in the soil. By keeping soil-inhabiting nematode populations in check, nematophagous fungi play a vital role in sub-soil ecosystems. Many *Cordyceps* species are also carnivorous, feeding on invertebrates.

The *decomposers* within the nutrient cycle of an ecosystem are the recyclers of waste and include bacteria, protozoans and fungi. In some respects, many invertebrates can play a role as decomposers too, as often they are the organism that does the mechanical work of breaking down the waste into smaller pieces.

The role of fungi as decomposers goes largely un-noticed as most of the work is done below the surface and we are only alerted to their presence by the seasonal appearance of their fruiting bodies.

Many of Australia's plant species are dependent on their specific fungal partners. These *mycoheterotrophic plants*, like our terrestrial orchids, acquire much or even all their nutrients through an association between their root system and the mycelium of the fungus.

**Fermentation** is yet another vital role within an ecosystem that requires a role by fungi – moulds and yeasts.

Organisms that live off other organisms are called *biotrophs*. Generally we are familiar with negative biotrophs that result in disease – viruses, bad gut microbes in animals, and rusts, smuts, mildews etc as plant pathogens – but there are many positive, or mutualistic, ones too; mycorrhiza and endophytic fungi are two of these.
Endophytes are present in many plant tissues and, although poorly understood, are thought to play a role in assisting plants to avoid disease and herbivore grazing, and some may sequester carbon.

Most terrestrial plants grow poorly, or not at all, without symbiotic fungi at their roots. Much of the carbohydrate produced by the plant during photosynthesis is passed down to the roots. Here, the fungal mycelia (‘roots’ of the fungi) exchange the carbohydrates from the plant for nitrogen, phosphorous and other nutrients obtained from the matter it has decomposed in the soil.

Most of the carbon passed to the fungi by the plant originally came from carbon dioxide in the atmosphere. Research now confirms that the majority of stored carbon in woodlands and forests is locked up in the plants’ root systems where it is used by mycorrhizal (‘root-associated’) fungi.

As well as being valuable carbon sinks, mycorrhizal fungi are thought to be one of the means by which plants have learnt to communicate with one another. Plants can secrete soluble chemicals into their root zones where fungi mycelia transport them to other nearby plants.

Mycophagy is the process of organisms consuming fungi. Many mammals, birds, insects, molluscs and other fungi often feed on fungi as part or all of their diet.

In conclusion, Dr McMullan-Fisher emphasised the need for consideration of the Fungi kingdom when issues of conserving biodiversity are proposed for a given ecosystem. In Australia, it is estimated that fungi represents around 9% of all known species and this is more than the vertebrates (2%) and the plants (5%) combined!

Dr McMullan-Fisher encouraged all present to report their fungi finds to Fungimap (fungimap.org.au). She also advised that the website has a steadily increasing gallery for identification purposes and many other tools to aid the amateur mycologist.

Flora, fauna AND fungi. Fungi are the ties that bind in terrestrial ecosystems.

Peter Ware

Excursion to Nyora 27.05.2017

This excursion to two reserves in Nyora was led by Jenny Rejske and Sapphire McMullan-Fisher. Jenny showed us where to go, and Sapphire told us what we were looking at and what it did in the ecosystem.

Our first reserve was the Henry Littledyke Reserve, which is also known as the Nyora Flora and Fauna Reserve and is 4 km east of Nyora. It was reserved partly because the rare and threatened Tea-Tree Fingers, which was first recorded nearby in Nyora in 1992, may have occurred there as it has a number of the host trees. Henry Littledyke, the owner of this property, was a community-minded pioneer who offered this site so a dam could be constructed to attract the railway to Nyora by providing water for the steam engines. The 24 ha site is a unique example of the diverse flora and fauna that covered the area prior to settlement.

Amazing coloured fungi did not jump out at us as we walked around, but nevertheless there was
no shortage of specimens – many of them fawn or brown, which seemed much harder to identify and get excited about.

*Fomitopsis lilacinogilva*, a shelf polypore, was growing on a tree stump. Growing on dead wood it causes a brown cubical rot which I realise I have often noticed on old wood. Its under-surface is lilac coloured, particularly on the margins, but like many of the specimens we saw it was past its prime and somewhat faded so I would have found it hard to identify from the fungi book.

Being very much a novice at identifying fungi, I’ve always found the sheer diversity quite overwhelming. When looking at the gilled fungi, I started to appreciate that features such as the cap shape (pointy, dished, rounded), texture (velvety, slimy) the length and thickness of the stipe, and whether it has a fringe, are all important things to consider. The spore colours are key identifying features, and if the time is right, that can be determined by looking at the spores that have fallen onto, and coloured, the stipe. Otherwise the specimen would need to be collected for its spore print.

Delicate, little *Marasmius* sp. were growing on a small stick. These grow in the leaf litter but regular burning will reduce their numbers. *Marasmius elegans*, a little red-capped toadstool, was more striking and had a distinctive two-toned stipe (white above and red-brown below). A cluster of short-stemmed, pointy-topped, light brown toadstools clustered on a dead branch was *Hypholoma fasciculare*. These may have lime-green gills when young that darken with age.

I learnt more about slime moulds with a growth of white ‘sponges on sticks’ seen on a log. These were the last stages of the fruiting bodies, and different to the stage I’m used to where they form a slimy mass.

*Mycena cystidiosa* was seen, including the hair-like rhizomorphs with minute capitate tips. I’ve seen these before, and they look like a mass of white hairs covering decaying matter. The fruiting bodies are a very tall, thin, greyish-brown toadstool with radial striations.

Eileen Laidlaw saw a small patch of *Favolaschia calocera* (Orange Ping-pong Bats) but deliberately did not point these out as they are a pest species and she didn’t want to risk spreading them. In hindsight she thought she should have made people aware of them so they would recognise them in future. It has spread around the world in recent times and occurs naturally in Asia and Madagascar. Cleaning shoes and other field clothing is an important part of protecting our bushlands.

After lunch we drove to the Wuchatsch Reserve, 5 km north west of Nyora. We got a lot more excited about the larger fungi we found here, especially when we got off the main track and were in the tea-tree where they were all around us in places.

Near the entrance were several *Descolea recedens*. These solid, golden-brown toadstools had the gill print still visible on what was left of the universal veil.

*Russula celandii* group were everywhere. However, most had been colonised by secondary growth of pin moulds (golden, hairy stems with a pin on top). There were also many boletes that were
being eaten by *Hypomyces chrysospermus* (the Bolete-eater). One was hosting a species of slime mould that was displaying the sexual stage, as was a *Stemonitis* species, which was very interesting and photographed by lots of people. It had a mass of erect, brown tubes rather like the bristles on a brush.

The entire side of a standing, live eucalypt was covered in very tiny *Dictyopanus pusillus* (Ping-Pong Bats). Sapphire noted they would be interesting to see at night time as this species is sometimes luminous.

Further up the track we discovered a *Fistulina hepatica* (Beefsteak Fungus). Sapphire removed this from the tree and cut it in half to expose the fertile tubes that extend all the way back through to the base of the fungi.

*Coltricia cinnamomea* looked particular beautiful with the brown, silken top that appeared to have radiating surface hairs and underneath had pores 1-3 mm in diameter. This species is a mycorrhizal partner of plants and loves heathland and sandy soil.

On the way home, Peter Ware took those of the group who remained to the Lang Lang Primary School Arboretum. This small block of land was formerly the school pine plantation, but has been revegetated and is now used by the school for environmental studies, some of which Peter has been leading. He wanted to show us some lovely specimens of the caterpillar fungus *Cordyceps gunnii*. There were quite a number growing under wattles, and the fresh fruiting specimens showed their striking lime-green stems and olive-green heads. This fungus parasitises a caterpillar (larvae of the moth genus *Oxycanus*). The spores are in the ground and penetrate the caterpillar when it pupates, consuming the soft tissue. The fruiting body that we see shoots up from the dead caterpillar.

This fungi excursion was very fruitful and informative, thanks to the abilities of Sapphire and Eileen to identify many of the fungi and tell us so much about what we saw. Thanks to Jenny too for leading us to all the right places.

Wendy Savage

*A list of fungi seen during this excursion is available in the electronic version of this Naturalist.*

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**Dean Ingwersen – Woodland Birds for Biodiversity**

On June 23rd, Dean Ingwersen spoke to the group about woodland bird projects at Birdlife Australia and, in particular, the recovery efforts he has been involved in coordinating for the endangered Regent Honeyeater.

Temperate woodlands used to be an extensive vegetation type in Tasmania and south-eastern Australia. These woodlands have over 250 species of birds living in them. One in four of these species is threatened or declining and some, including the Regent Honeyeater, Swift Parrot and Grey-crowned Babbler are now thought to be in danger of extinction. In NSW, Gang-gang Cockatoo populations are also declining.

The area of available habitat for woodland birds has greatly contracted since European settlement. To date, 85% percent of temperate woodlands have been cleared for agriculture. The majority of remaining areas suitable as habitat are along rivers or roads and on private property.

Dean’s recent work centres on species recovery for the Regent Honeyeater, and the second part of
his talk was mainly about this species. What is known is that it used to be very common, with flocks of 1000 or more birds reported. It is now extinct in South Australia. There are a few left in western Victoria. The total Australian population is thought to be 1500 birds or fewer, of which less than 100 are in Victoria, and 65% of records of Regent Honeyeater sightings have been from private property.

Birdlife Australia’s efforts in conservation have involved research to predict likely areas of habitat, approaching landholders to assess their interest and then putting covenants on areas that can be preserved in perpetuity as habitat.

There are also other community-led projects that aim to restore habitat by planting suitable species and fencing remnant vegetation. Over 30,000 people have been involved in one planting project around Benalla in northern Victoria [http://regenthoneyeater.org.au/](http://regenthoneyeater.org.au/).

Work at Birdlife Australia on species conservation has been challenged by changes in federal and state government funding regimes, but there is now a ten-year conservation contract process in NSW which has provided some certainty, so projects in that state have been able to be prioritised.

Dean has been involved in banding, monitoring and managing known populations of birds. There is a captive breeding program for Regent Honeyeaters at Taronga Zoo and other locations. One of the release sites for the captive-bred birds is at Chiltern, where our group will be going for the spring camp. Dean expressed his hope that he will be able to meet us when we’re there and show us some Regent Honeyeater breeding sites.

One bird that has thrived under widespread land-clearing has been the Noisy Miner. This communal-breeding and extremely territorial honeyeater flourishes at the expense of many other species. Noisy Miners attack other vulnerable species and pull apart their nests. Although they are native to Australia, they are now listed as a “key threatening process.” Part of Regent Honeyeater conservation in the Kapertee Valley in the Hunter Region of NSW has been to try and create a buffer around known Regent Honeyeater breeding sites by controlling Noisy Miner numbers. This was undertaken by employing professional expert shooters to kill large numbers of miners (which were quickly replaced in the same areas by slightly lower numbers of the same species). Much effort went into getting the correct permits for this program. This part of Dean’s talk proved to be a bit distressing for some in the audience and was of great interest to others who would also like to reduce the numbers of this bird in some local areas to allow other smaller birds to stand a chance.

Although they have shown considerable success in long-distance flying, the captive-bred birds that are being monitored have yet to breed successfully in the wild. Efforts to find out the causes involved 24-hour monitoring of some nests with night-vision cameras. Those in our group involved with providing nesting boxes for Sugar Gliders may have been appalled to see their little furry friends exposed as a nocturnal nest predator, as was the cute and endangered Squirrel Glider, and, at another site, brush-tailed possums. Known diurnal predators include currawongs and raptors.

Some birds have been found breeding and feeding well outside of their known range and away from what is thought to be their preferred habitat. Dean is about ready to say “any flowering tree” is where they may be found. He handed out some useful identification guides that help to distinguish these birds from other yellow-winged honeyeaters.

In a footnote to this interesting and well-attended talk, we were asked to remember a ‘Birds on
Twelve of us ascended the hills south of Yinnar South in three four-wheel drive vehicles. We walked down the vehicle track HVP32, which was bordered with prolific regrowth of shrubs such as Cassina longifolia, Goodenia ovata, Olearia lirata, Senecio linearifolia and Cassinia aculeata. The air was fresh and cool, pleasantly scented by the forest plants. The weather was mostly fine with periods of sunshine and occasional light showers.

We took a partly overgrown track through a previously logged area. Jay explained that, following clear-felling about 8 years ago, HVP has replanted, as required, before returning this block to public ownership. A few older trees of some girth stood here and there along the track: Mountain Ash Eucalyptus regnans, Mountain Grey Gum Eucalyptus cypellocarpa and Blackwood Acacia melanoxylon. Regrowth included Silver Wattle, Varnish Wattle, Musk Daisy-bush, Hazel Pomaderris, Austral Mulberry, Bootlace Bush and Mountain Pepper. Smaller plants beside the track included Common Ground-fern, Rough Tree-fern, Bracken, Tall Sedge, Forest Wiregrass, Prickly Currant-bush, Geranium, Hairy Pennywort, Ivy-leaf Violet, Forest Starwort, and a scattering of a variety of small weeds. Ken pointed out Trailing St John’s Wort, a small prostrate weed with tiny black spots on the margins of the leaves.

We drove to a clearing further along HVP32. Here we were outside the section of College Creek that was returned to the public. We were entertained by Grey Fantails as we lunched. We admired an unusually large Purple Apple-berry Billardiera macrantha climbing high on a tree trunk and hung with a few bright, shiny, purple berries. We were to see more of this plant on the next track. From the clearing we chose to follow a very overgrown track, where in places we had to push through wet vegetation, to explore the south-facing slope. We added a variety of ferns to our list: Fishbone Water-fern, Ray Water-fern, Hard Water-fern, Bat’s-wing Fern, Mother Shield-fern and Scrambling Coral-fern. Tree Zieria, also known as Stinkwood, created some interest when the reason for one of its common names was experienced. It was in bud.


There is now some new funding available to revive this program and to monitor previous locations. The catch is that the locations were supplied to Dean as latitude and longitude without addresses or landholder names. If you can help, Dean’s contact details are:

Dean Ingwersen – Project Manager & Regent Honeyeater Recovery Coordinator
T 1800 621 056
E dean.ingwersen@birdlife.org.au

Jay Duncan
Several tiny soil fungi added colour, and a Split-gill decorated a decaying log.

Back at the clearing, after some hesitation – perhaps due to another brief shower of rain – we walked further along HVP32. As we set off, several types of bracket fungi on decaying logs attracted our attention. We passed a stand of young eucalypts on the lower side of the track. These were probably a plantation for future logging. We confirmed the presence of both *Clematis aristata* and *C. glycinoides*. Along the sides of the track we found Blanket Bush, Banyalla, Shade Nettle, Scrub Nettle, Hemp Bush, *Dianella tasmanica*, Forest Hound’s-tongue, Shade Plantain, Kangaroo Fern, and a wide range of small, common weeds.

Margaret Rowe

*A plant list for this excursion is available in the electronic version of this Naturalist.*

### Moe-Yallourn Rail Trail excursion 29.07.2017

In 1997, on the 6th and 29th of September, and the 15th of November, the Latrobe Valley Field Naturalists Club conducted floral and faunal surveys of the Moe to Yallourn rail easement. The data gathered was used in the *Moe to Yallourn Rail Trail Draft Concept and Business Plan* in February 1998. LVFNC member Madge Vinnell was a member of the Management Committee. Again, after confirmation that “the information contained in them is still relevant at the present time”, this data was used in the Plan’s update in March 2009. In 2012, Indigenous Design was tasked with developing a management plan for the Committee of Management.

Our excursion was planned and led by Alix Williams and Margaret Coupe. Margaret is the Secretary of the Friends of the Moe-Yallourn Rail Trail and she had asked for a further survey and suggestions for future planting along the Trail. We met at the Moe Botanic Gardens carpark to car pool as a Sea-eagle flew over – a good start to a sunny winter’s day. We drove to the Yallourn Power Station and parked at the entrance to the Rail Trail. Time was spent there beginning the botany list and then looking at the spring that had appeared some years ago and caused damage to the trail, necessitating major drainage works and a retaining wall. A Common Eastern Froglet was heard expressing appreciation of the area.

We drove on to Pettits Track and walked over the bridge to the south where we looked down into a valley leading to the power station. The vegetation here is highly-rated Lowland Forest. Along the track, Sydney Wattles added a splash of gold and Yellow Robins and Kookaburras called. Further on we stopped to look down a very steep slope to Lake Narracan where we heard a Darter calling and saw Superb Fairy-wrens and Brown Thornbills, and a Straw-necked Ibis flew over. Margaret Coupe explained the work that had been done to clear the nearby area of Blackberries, Burgan and other weeds, and showed us the area that had been recently planted with indigenous trees and shrubs. Further along the Trail, Margaret pointed out a slope where Radiata Pines had been taken out to open up a very dark area and she asked us to provide suggestions for replanting the steep slope.

Margaret expressed appreciation for the work done by the Lowanna Secondary College Year 9 students over the past 10 years, giving over 250 hours of work along the Trail, pulling weeds, painting signs and removing Sweet Pittosporum. Also, Baringa Special School students had planted 160 tree seedlings last week.

We stopped again at the pedestrian bridge over the Narracan Creek and walked back to see a stand of the rare Strzelecki Gums.

At the end of the Trail, Margaret invited us to contribute ideas regarding the design of a proposed
information board for the beginning of the Trail. It was a most interesting excursion and appreciation was expressed to Margaret and the members of the Friends of the Moe-Yallourn Rail Trail for their dedication and hard work in keeping the Trail open and in good condition for use by the community and the preservation of native flora and fauna.

After lunch at the Botanic Gardens we drove to Sullivans Track and down to Lake Narracan where we were successful in seeing some more birds.

Thanks again to Margaret and Alix for a most informative and well-planned excursion.

Meryl Cracknell

*A bird list for this excursion is available in the electronic version of this Naturalist.*

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**Birdlife Australia Challenge Count 2017**

As our December “excursion”, members of the LVFNC, their relatives and friends take part in the Australia–wide Bird Challenge Count. We have been doing this since 1998. Participants join groups that visit a series of sites in order to observe and count birds. Some people are able to identify birds and others assist by spotting and counting them. Each group starts around 8.30 am and finishes mid-late afternoon. Data on the types and numbers of birds recorded are sent to Birdlife Australia. This information provides ideas about bird populations and movement in this area.

- **Group 1** – Thursday Nov 30: Energy Australia Wetlands, Crinigan Road Reserve
- **Group 2** – Friday Dec 1: Edward Hunter, Moe Treatment Works, Lake Narracan
- **Group 3** – Saturday Dec 2: Yarragon South, Uralla Reserve, Trafalgar Settlement Ponds
- **Group 4** – Saturday Dec 2: Traralgon Railway Reservoir Conservation Reserve, Wirilda
- **Group 5** – Sunday Dec 3: Mathison Park, Morwell NP
- **Group 6** – Sunday Dec 3: Jeeralang Junction

If you would like to join in the fun (for part or all of a day) and haven’t yet contacted Alix, please email alixw@bigpond.com or phone 5127 3393.

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**WELCOME TO NEW MEMBERS**

The Club welcomes Adrian Clements from Traralgon. We wish you a long and happy association with us.

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**CHRISTMAS PARTY**

The Club’s Christmas Party this year will be held at John & Jan Tulloch’s place at 55 Lynne Avenue, Moe South on Saturday 9th December. At Moe, from the roundabout over the rail line, head out High St to Moe South. Follow the white line through road junctions for 4.9km (it will have become Moe South Rd) until you reach Lynne Avenue on your right. The Tullochs are 550m along on the left. Celebrations commence at 12 noon. Please BYO food, drinks, table and chairs. Punch, Christmas cake, tea and coffee provided, and of course... the Quiz!
REPORT ON BUSINESS MEETING 23.10.2017

Finance

Cash Management Trading Account: $4,908.34  Term Deposit: $17,032.88

Business Arising, Correspondence & General Business

- Spending of club funds: Signage at Dawson Rail Trail is falling into disrepair, so Ken Smith suggested we contact Parks Victoria to offer to assist with funding. Wendy will contact Ken and make investigations.

- Digitising LVFNC and Bon & Ollie Thompson’s plant lists: Phil has spoken to Eric Eklund and he is keen to support the project but wants us to seek funding to cover the digitisation and storage preparation.

- Club spring camp at Chiltern: Very successful with 27 participants. Going to send *Gippsland Lady Botanists* book to Eileen Collins. Ken sent photos of moths and other insects to Eileen and they will go on the Friends of Chiltern-Mt Pilot NP website.

- SEANA spring camp at Little Desert: had a record attendance of 135 including 12 of our club members. Club purchased *Birds and Plants of the Little Desert* book for our members to borrow.

- Purple Diuris count: Dawson has been burnt so very few orchids, will not do count. Longford Rd should be done this week.

- Latrobe Valley Mine Rehabilitation Consultative Group looking for LVFNC representative: David Stickney is already on the Hazelwood group and will follow up with Alix.

- ABN Register update: Phil has updated this and will remain the registered contact, adding David Mules as a second contact.

- National Disability Insurance Scheme is assessing suitability of local interest groups for people with a disability, and requested information about our activities. Will reply with details.

- Club summer camp at Mt Buffalo Fri 2 – Tues 6 February 2018. Accommodation at Porepunkah Bridge Caravan Park on the Mt Buffalo Rd have a good number of cabins and camping available. Can get a discount package with an estimate of numbers. People should book their own cabins as for the Chiltern camp.

Conservation matters

- Platypus drowning in yabby nets: Further response from Assistant Minister for Agriculture, Anne Ruston. Will respond and ask for details of their enforcement and education programs.

- VicForests proposed logging at Mirboo North: Phil sent a letter from the Club. Reply from Lily D’Ambrosio unsatisfactory. Contact from sub-group re biodiversity aspects, wanting advice on mammal and owl surveys. Rolf Willig is involved, and Ken Smith has prepared a more comprehensive orchid list.

- Replacement of bridge on Traralgon-Tyers Rd: Club has put in an objection and we should be advised about when it will be discussed by Council, possibly Nov 13 meeting. David Stickney will attend to support Irene.

- DELWP forest regulation: opportunity to respond. Wendy will circulate and people can send ideas to Phil who will respond if need be.

- Cores and Links handback: details have been sent and it is moving in the right direction.

- Jackie is concerned about the push for development of onshore gas extraction to be revisited, and will prepare information about the dangers of fracking for people to view at the general meeting.
Guest speaker for November

Martin O’Brien

Martin is a wildlife biologist who worked for 25 years as a policy officer with the Department of Environment. He will speak about various aspects of the Strzelecki Ranges warm temperate rainforest.

Summer Members’ Night

Various speakers

January’s meeting is our annual Summer Members’ Night, where club members can present a short talk (~10 mins) on any topic related to natural history. A projector is available for showing Power Point presentations or photos, if required. If you would like to present to the group on this night, please register your interest by emailing David Stickney at david.stickney@westnet.com.au.

Thank you to everyone who contributed to The LV Naturalist in 2017. Best wishes for a Merry Christmas and a safe and happy New Year to all our members, their families and friends.

Deadline for articles to be considered for inclusion in the next issue (Jan/Feb): 8 Jan 2018

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Latrobe Valley Naturalist
Issue no. 593
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APPENDICES

APPENDIX I – Fungi list for Nyora excursion 27.05.2017 (W. Savage)

Henry Littledyke Reserve

Bolbitius vitellinus  
Cruentomycena viscidocruenta  
Favolaschia calocera*  
Fomitopsis lilacinogilva  
Galerina sp.  
Hypholoma fasciculare  
Macrolepiota clelandii

Marasmius elegans  
Marasmius sp.  
Marasmius alveolaris  
Mycena cystidiosa  
Mycena vinacea  
Oudemansiella gigaspora group  
Phellinus sp.

Wuchatsch Reserve

Amanita sp.  
Coltricia cinnamomea  
Cortinarius sinapicolor  
Cortinarius sp  
Crepidotus crocophyllus  
Descolea recedens  
Dictyoandon pusillus  
Discinella terrestris  
Fistulina hepatica  
Gymnopilus allantopus

Hypomyces chrysospermus  
Lactarius eucalypti  
Melanoleuca sp.  
Mycena australororida  
Panellus pusillus  
Russula clelandii group  
Russula sp.  
Stemonitis sp.  
Corticioid fungi  
Stereum ostrea

Lang Lang Primary School Arboretum

Cordiceps gunnii  
Pycnoporus sp.

Geastrum triplex  
*introduced
## APPENDIX II – Plant list for College Creek excursion 24.06.2017 (M. Rowe)

### Ferns

<table>
<thead>
<tr>
<th>Family</th>
<th>Species</th>
<th>Common Name</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blechnaceae</td>
<td>Blechnum fluitiate</td>
<td>Ray Water-fern</td>
</tr>
<tr>
<td>Blechnaceae</td>
<td>Blechnum nudum</td>
<td>Fishbone Water-fern</td>
</tr>
<tr>
<td>Blechnaceae</td>
<td>Blechnum wattsii</td>
<td>Hard Water-fern</td>
</tr>
<tr>
<td>Cyatheaceae</td>
<td>Cyathea australis</td>
<td>Rough Treefern</td>
</tr>
<tr>
<td>Dennstaedtiaceae</td>
<td>Histiopteris incisa</td>
<td>Bat's-wing Fern</td>
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<td>Pteridium esculentum</td>
<td>Austral Bracken</td>
</tr>
<tr>
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<td>Calochlaena dubia</td>
<td>Common Ground-fern</td>
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<tr>
<td>Dryopteridaceae</td>
<td>Polystichum proliferum</td>
<td>Mother Shield-fern</td>
</tr>
<tr>
<td>Gleicheniaceae</td>
<td>Gleichenia microphylla</td>
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<td>Polypodiaceae</td>
<td>Microsorum pustulatum</td>
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### Monocotyledons

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<thead>
<tr>
<th>Family</th>
<th>Species</th>
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<tbody>
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<td>Carex appressa</td>
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<td>Juncaceae</td>
<td>Juncus sp.</td>
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<td>Juncaceae</td>
<td>Luzula meridionalis</td>
<td>Field Woodrush</td>
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<td>Liliaceae</td>
<td>Dianella tasmanica</td>
<td>Tasman Flax-lily</td>
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<td>Poaceae</td>
<td>Tetrarrhena juncea</td>
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### Dicotyledons

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<tr>
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<tbody>
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<td>Hairy Pennywort</td>
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<td>Bedfordia arborescens</td>
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<td>Asteraceae</td>
<td>Cassinia aculeata</td>
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<td>Cassinia longifolia</td>
<td>Shiny Cassinia</td>
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<tr>
<td>Asteraceae</td>
<td>Cirsium vulgare*</td>
<td>Spear Thistle</td>
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<tr>
<td>Asteraceae</td>
<td>Conyza sp. *</td>
<td>Fleabane</td>
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<td>Cotula australis</td>
<td>Common Cotula</td>
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<td>Cudweed</td>
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<td>Cat's-ear</td>
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<td>Asteraceae</td>
<td>Leucanthemum vulgare*</td>
<td>Oxeye Daisy</td>
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<td>Asteraceae</td>
<td>Olearia argophylla</td>
<td>Musk Daisy-bush</td>
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<td>Snowy Daisy-bush</td>
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<td>Olearia phlogopappa</td>
<td>Dusty Daisy-bush</td>
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<tr>
<td>Asteraceae</td>
<td>Senecio jacobaea*</td>
<td>Ragwort</td>
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<td>Senecio linearifolius</td>
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<td>Senecio minimus</td>
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<td>Austrocynoglossum latifolium</td>
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<td>Hypericum humifusum*</td>
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<td>Mentha pulegium*</td>
<td>Pennyroyal</td>
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<td>Prostanthera laeianthos</td>
<td>Christmas Bush</td>
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<td>Lamiaceae</td>
<td>Prunella vulgaris*</td>
<td>Self-heal</td>
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<td>Gynatrix pulchella</td>
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<td>Eucalyptus cypellocarpa</td>
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<td>Plantago coronopus*</td>
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<td>Solanum furcatum*</td>
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<td>Tasmannia lanceolata</td>
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*introduced
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<tr>
<td>Australasian Darter</td>
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<td>Red Wattlebird</td>
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<td>Straw-necked Ibis</td>
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<td>Superb Fairy-wren</td>
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<tr>
<td>Eastern Yellow Robin</td>
<td>Welcome Swallow</td>
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<tr>
<td>Laughing Kookaburra</td>
<td>White-bellied Sea-eagle</td>
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<td>Little Raven</td>
<td>White-eared Honeyeater</td>
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